Biology 265 Bevington
Cell Physiology Spring Term, 2012

COURSE SYLLABUS

Text: Alberts, B. et al. 2008. Molecular Biology of the Cell. Garland Publishing Co., New York. Fifth Edition.

Course Objectives:

The course will examine the structural elements of cells and their physiological functions. Both procaryotic and eucaryotic cells will be studied, but the emphasis will be on eucaryotic ones. Molecular interactions between complex molecules will be studied as a way to understand the cellular relationships between structure and function.

Considerable attention will be given to cell membranes, especially to their roles in transport phenomena, cell-to-cell signaling, and protein targeting processes. Basic features of intermediate metabolism and metabolic control will be covered, particularly those pathways which cells use to derive energy. Genetic mechanisms to be studied include the basic elements of gene structure and control of gene expression in procaryotes and eucaryotes. The cell cycle and its control will be examined, and some time will be devoted to cancer cells where control of the cell cycle has been lost.

The laboratory will emphasize techniques and methodology in cell biology. Since the accurate interpretation of data presupposes a firm understanding of how the data were acquired, it is hoped that you will develop an interest in the techniques and procedures we use in the laboratory. You should understand not only the theory behind the experiments but also principles underlying the protocols. In science, the methods one uses are as important as the results one observes.

Prerequisites:

Chemistry 113 and 114; Biology 112 and/or 119.

About the text:

Reading assignments will cover about half of the textbook. In addition to its use in this course the text should be a good reference for other biology courses. From time to time the instructor will provide handouts and copies of recent journal articles to supplement certain lecture topics. These will be principally in electronic form. Handouts and especially the articles from the literature should be considered as important as the reading assignments from the text.

A separate handout with all of the semester reading assignments in the text will be distributed on the first day of class. You are encouraged to use the DVD that comes with the text throughout the semester. It includes short videos, animations, and a variety of learning tools.

Grading:

Item	Point Value	Percentage of Final Grade
Three hour exams (180 points each)	540	54%
Three lab quizzes (65 points each)	195	19.5%
One final exam (comprehensive)	265	26.5%
	1000	100%

Exams and quizzes are scheduled through the semester so that there will be only one of either of them in any given week.

Grading Scale:

90 - 100	A
80 - 89	B
70 - 79	C
60 - 69	D
< 60	F

LECTURE SEQUENCE[1]

Introduction: scope and objectives of the course

A brief review of cell structure

Fractionation of cellular organelles

Molecules in cells

- Chemical bonds and molecular interactions
- Important characteristics of water
- The major classes of small molecules (sugars, amino acids, fatty acids, and nucleotides)
- Nucleic acids
- Proteins
  - Structure
  - Protein functions and factors which affect function
  - Assembly and turnover
- Enzymes: kinetics and factors affecting catalytic action
- Lipids and carbohydrates*

How cells synthesize proteins

- Basic mechanisms of the process
- RNA synthesis, RNA processing, RNA export from the nucleus
- Molecular chaperones and protein folding
- Ubiquitin, proteasomes, and protein destruction
- Transcriptional control of gene expression
- Posttranscriptional control

Cell growth and control of the cell cycle

The mechanics of cell division*

Cancer cells and loss of control of the cell cycle

Cellular membranes

- Architecture

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[1] A few of the lecture topics marked with an asterisk (*) may be abbreviated or eliminated to allow more time for other topics.

Cell junctions*
Transport phenomena

Cellular compartmentalization and protein sorting:

How are proteins targeted to and from the nucleus, mitochondria, chloroplasts, and peroxisomes?

The endoplasmic reticulum

Signal hypothesis and the role of SRPs in directing proteins to the ER
Vectorial transport of proteins into the ER lumen
Protein glycosylation
How the ER deals with improperly folded proteins
Synthesis of membrane lipids

Vesicular traffic: How do vesicles arrive at the correct destinations within the cell?

The Golgi complex

Structure and origin
Posttranslational modification of secretory, membrane, and glycoproteins
Sorting, packaging, and targeting of proteins from the Golgi

Lysosomes and cellular digestion

Receptor-mediated endocytosis

Exocytosis and secretion

Cell signaling

General principles
Signaling via G-protein linked cell surface receptors

Cytoskeleton*

Nature of the cytoskeleton
Motor proteins
Cilia and flagella

Cell metabolism

Overview and general concepts
The major metabolic pathways of cells

How cells regulate metabolism

Glyolysis in the cytoplasm

The mitochondrion

Oxidation of pyruvate and fatty acids in the citric acid cycle
Electron transport, proton-motive force, and oxidative phosphorylation

Metabolic regulation of glycolysis and citric acid cycle

The Glyoxylate cycle and lipid metabolism

The pentose shunt and why cells have it

The chloroplast and photosynthesis

Photochemical events and the light reactions
Photolysis, electron transport, and photophosphorylation
C_3 and C_4 metabolism

TARGET DATES FOR LECTURE TOPICS

Mon.	16 Jan.	Introduction, course objectives, begin review of cell structure
Wed.	18 Jan.	Review of cell structure
Fri.	20 Jan.	Complete review of cell structure; fractionation of cellular organelles; begin chemical bonds and molecular interactions
Mon.	23 Jan.	Properties of water, small molecules
Wed.	25 Jan	Nucleic acids
Fri.	27 Jan.	Proteins
Mon.	30 Jan.	Proteins
Wed.	1 Feb.	**1st Quiz** (labs 1 & 2); proteins
Fri.	3 Feb.	Enzymes
Mon.	6 Feb.	Enzymes
Wed.	8 Feb.	Protein synthesis
Fri.	10 Feb.	**FIRST HOUR EXAM**
Mon.	13 Feb.	Protein synthesis
Wed.	15 Feb.	Protein synthesis
Fri.	17 Feb.	Control of gene expression
Mon.	20 Feb.	Control of gene expression
Wed.	22 Feb.	Control of gene expression
Fri.	24 Feb.	Cell cycle (MID TERM)
Mon.	27 Feb.	**2nd Quiz** (labs 3, 4, 5 & 6); cell cycle
Wed.	29 Feb.	Cancer cells
Fri.	2 Mar.	Cancer cells; begin cell membranes
Sat.	3 Mar. - Sun. 11 Mar.	**SPRING RECESS**
Mon.	12 Mar.	Cellular membranes
Wed.	14 Mar.	Cellular membranes
Fri.	16 Mar.	Targeting proteins to the nucleus, mitochondria, and chloroplasts
Mon.	19 Mar.	**SECOND HOUR EXAM**

Wed.	21 Mar.	Endoplasmic reticulum
Fri.	23 Mar.	ER and Golgi
Mon.	26 Mar.	Golgi
Wed.	28 Mar.	Golgi
Fri.	30 Mar.	Lysosomes; receptor-mediated endocytosis
Mon.	2 Apr.	Complete lysosomes
Wed.	4 Apr.	Cell signaling
Fri.	6 Apr. - Mon. 9 Apr.	**EASTER RECESS**
Wed.	11 Apr.	**3rd Quiz** (labs 7, 8, and 9); cell signaling
Fri.	13 Apr.	Overview of metabolism, glycolysis
Mon.	16 Apr.	Glycolysis
Wed.	18 Apr.	Glycolysis
Fri.	20 Apr.	TCA cycle
Mon.	23 Apr.	TCA cycle and its control, cytochrome system, oxidative phosphorylation
Wed.	25 Apr.	**THIRD HOUR EXAM**
Fr..	27 Apr.	Pentose phosphate pathway and the glyoxylate cycle
Wed.	28 Apr.	Review and comparisons: respiratory metabolism and photosynthesis
Fri.	30 Apr.	Photosynthesis
Mon.	30 Apr - Fri. 4 May	Final Exam Period
Fri.	4 May 8:30 am	Final exam date for the course

LABORATORY SCHEDULE

Lab. No.	Dates	Topics
1.	17, 18 Jan.	Laboratory orientation: protocols, preparing solutions and making dilutions, pipeting, pipeting devices, and safety precautions
2.	24, 25 Jan.	Spectrophotometry: Beer's Law, use of the B&L Spectronic 20 spectrophotometers Constructing absorption curve for DCPIP and anthocyanin pigments at different pH values
3.	31 Jan. 1 Feb.	Spectrophotometric assays for protein (Bradford and bicinchoninic acid methods)
4.	7, 8 Feb.	Cell growth: growth kinetics in *Enterobacter aerogenes* 1. Determining generation time 2. Effects of temperature, chloramphenicol, peptone, and an amino acid analogue
5.	14, 15 Feb.	Enzyme assay: acid phosphatase 1. Effect of substrate concentration 2. Effect of phosphate ion
6.	21, 22 Feb.	Estimation of specific activity of extracted acid phosphatase
7.	28, 29 Feb.	Isolating an organelle: 1. Mitochondria from cauliflower florets 2. Enzyme assay for succinic dehydrogenase and/or malate dehydrogenase
	Sat. 3 Mar. - Sun. 11 Mar.	**Spring Recess**
8.	13, 14 Mar.	Factors affecting membrane permeability

9. 20, 21 Mar. Tyrosinase/phenol oxidase

10. 27, 28 Mar. Hill reaction and the Emerson enhancement effect in isolated chloroplasts

11. 3, 4 Apr. Manometric measurement of respiratory activity using the Gilson respirometer
    1. Effects of temperature, substrates, and inhibitors on yeast cell suspensions (*Saccharomyces cerevisiae*)
    2. Effects of washing and aging in discs of storage tissue from potato tubers (*Solanum tuberosum*)

Fri. 6 Apr. - Mon. 9 Apr**.** **Easter Recess**

12. 10, 11 Apr. Effects of ultraviolet radiation on *Paramecium*, *Tetrahymena*, and *Euglena*

13. 17, 18 Apr. Open

14. 24, 25 Apr. Effects of cycloheximide and colchicine on flagella regeneration in *Chlamydomonas reinhardi* (or) review

SUMMARY OF DEADLINES

Wed.	1 Feb.	First lab quiz (No's. 1 and 2)
Fri.	10 Feb.	FIRST HOUR EXAM
Mon.	27 Feb.	Second lab quiz (No's. 3, 4, 5 and 6)
Mon.	19 Mar.	SECOND HOUR EXAM

Wed.	11 Apr.	Third lab quiz (No's 6, 7, 8 and 9)
Wed.	25 Apr.	THIRD HOUR EXAM
Fri.	4 May	FINAL EXAM (8:30 am)

Learning Disability Statement *– Students who wish to request accommodations in this class for a disability should contact Mr. Joe Kempfer, Assistant Director of Learning Services for Disability Support, 1307 Main Street (extension 1510). Accommodations cannot be provided until authorization is received from the office of Learning Services.*