

COURSE SYLLABUS

Text: Alberts, B. et al. 2008. *Molecular Biology of the Cell*. Garland Publishing Co., New York. Fifth Edition.

Course Objectives:

The course will examine the structural elements of cells and their physiological functions. Both procaryotic and eucaryotic cells will be studied, but the emphasis will be on eucaryotic cells. Molecular interactions between complex molecules will be studied as a way to understand the cellular relationships between structure and function.

Considerable attention will be given to cell membranes, especially their roles in transport phenomena, cell-to-cell signaling, and protein targeting processes. Basic features of intermediate metabolism and metabolic control will be covered, particularly those pathways which cells use to derive energy. Genetic mechanisms to be studied include the basic elements of gene structure and control of gene expression in procaryotes and eucaryotes. The cell cycle and its control will be examined, and some time will be devoted to cancer cells where control of the cell cycle has been lost.

The laboratory will emphasize techniques and methodology in cell biology. Since the accurate interpretation of data presupposes a firm understanding of how the data were acquired, it is hoped that you will develop an interest in the techniques and procedures we use in the laboratory. You should understand not only the theory behind the experiments but also principles underlying the protocols. In science, the methods one uses are as important as the results one observes.

Prerequisites:

Chemistry 113 and 114; Biology 112 and/or 119.

About the text:

Reading assignments will cover about half of the textbook. In addition to its use in this course the text should be a good reference for other biology courses. From time to time the instructor will provide handouts and copies of recent journal articles to supplement certain lecture topics. These may be either in paper or electronic form. Handouts and especially the articles from the literature should be considered as important as the reading assignments from the text.

Grading:

<u>Item</u>	<u>Point Value</u>	<u>Percentage of Final Grade</u>
Three hour exams (180 points each)	540	54%
Three lab quizzes (65 points each)	195	19.5%
One final exam (comprehensive)	265	26.5%
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	1000	100%

Exams and quizzes are scheduled through the semester so that there will be only one of them in any given week.

LECTURE SEQUENCE¹

Introduction: scope and objectives of the course

A brief review of cell structure

Fractionation of cellular organelles

Molecules in cells

Chemical bonds and molecular interactions

Important characteristics of water

The major classes of small molecules (sugars, amino acids, fatty acids, and nucleotides)

Nucleic acids

Proteins

Structure

Protein functions and factors which affect function

Assembly and turnover

Enzymes: kinetics and factors affecting catalytic action

Lipids and carbohydrates*

How cells synthesize proteins

Basic mechanisms of the process

RNA synthesis, RNA processing, RNA export from the nucleus

Molecular chaperones and protein folding

Ubiquitin, proteasomes, and protein destruction

Transcriptional control of gene expression

Posttranscriptional control

Cell growth and control of the cell cycle

The mechanics of cell division*

Cancer cells and loss of control of the cell cycle

Cellular membranes

Architecture

Cell junctions*

¹ A few of the lecture topics marked with an asterisk (*) may be abbreviated or eliminated to allow more time for other topics.

Transport phenomena

Cellular compartmentalization and protein sorting:

How are proteins targeted to and from the nucleus, mitochondria, chloroplasts, and peroxisomes?

The endoplasmic reticulum

Signal hypothesis and the role of SRPs in directing proteins to the ER

Vectorial transport of proteins into the ER lumen

Protein glycosylation

How the ER deals with improperly folded proteins

Synthesis of membrane lipids

Vesicular traffic: How do vesicles arrive at the correct destinations within the cell?

The Golgi complex

Structure and origin

Posttranslational modification of secretory, membrane, and glycoproteins

Sorting, packaging, and targeting of proteins from the Golgi

Lysosomes and cellular digestion

Receptor-mediated endocytosis

Exocytosis and secretion

Cell signaling

General principles

Signaling via G-protein linked cell surface receptors

Cytoskeleton *

Nature of the cytoskeleton

Motor proteins

Cilia and flagella

Cell metabolism

Overview and general concepts

The major metabolic pathways of cells

How cells regulate metabolism

Glycolysis in the cytoplasm

The mitochondrion

Oxidation of pyruvate and fatty acids in the citric acid cycle
Electron transport, proton-motive force, and oxidative phosphorylation

Metabolic regulation of glycolysis and citric acid cycle

The Glyoxylate cycle and lipid metabolism

Pentose shunt

The chloroplast and photosynthesis

Photochemical events and the light reactions
Photolysis, electron transport, and photophosphorylation
C₃ and C₄ metabolism

TARGET DATES FOR LECTURE TOPICS

Mon.	17 Jan.	Introduction, course objectives, begin review of cell structure
Wed.	19 Jan.	Review of cell structure
Fri.	21 Jan.	Complete review of cell structure; fractionation of cellular organelles; begin chemical bonds and molecular interactions
Mon.	24 Jan.	Properties of water, small molecules
Wed.	26 Jan.	Nucleic acids
Fri.	28 Jan.	Proteins
Mon.	31 Jan.	Proteins
Wed.	2 Feb.	1st Quiz (labs 1 & 2); proteins
Fri.	4 Feb.	Enzymes
Mon.	7 Feb.	Enzymes
Wed.	9 Feb.	Protein synthesis
Fri.	11 Feb.	FIRST HOUR EXAM
Mon.	14 Feb.	Protein synthesis
Wed.	16 Feb.	Protein synthesis
Fri.	18 Feb.	Control of gene expression
Mon.	21 Feb.	Control of gene expression
Wed.	23 Feb.	Control of gene expression
Fri.	25 Feb.	Cell cycle
Mon.	28 Feb.	Cell cycle
Wed.	2 Mar.	2nd Quiz (labs 3,4, 5 and 6); start cancer cells
Fri.	4 Mar.	Cancer cells; begin cell membranes (MID TERM)
Sat.	5 Mar. - Sun.	13 Mar. SPRING RECESS
Mon.	14 Mar.	Cellular membranes
Wed.	16 Mar.	Cellular membranes
Fri.	18 Mar.	Targeting proteins to the nucleus, mitochondria, and chloroplasts
Mon.	21 Mar.	SECOND HOUR EXAM
Wed.	23 Mar.	Endoplasmic reticulum

Fri.	25 Mar.	ER and Golgi
Mon.	28 Mar.	Golgi
Wed.	30 Mar.	Golgi
Fri.	1 Apr.	Lysosomes; receptor-mediated endocytosis
Mon.	4 Apr.	Complete lysosomes
Wed.	6 Apr.	3rd Quiz (labs 7, 8, 9 and 10), Cell signaling
Fri.	8 Apr.	An overview of metabolism, glycolysis
Mon.	11 Apr.	Glycolysis
Wed.	13 Apr.	Glycolysis
Fri.	15 Apr.	TCA cycle
Mon.	18 Apr.	TCA cycle and its control, cytochrome system, oxidative phosphorylation
Wed.	20 Apr.	THIRD HOUR EXAM
Fri.	22 Apr. - Mon. 25 Apr.	EASTER RECESS
Wed.	27 Apr.	Pentose phosphate pathway and the glyoxylate cycle
Fri.	29 Apr.	Review and comparisons: respiratory metabolism and photosynthesis (Last day of classes)
Mon.	2 May - Sat. 7 May	Final Exam Period

LABORATORY SCHEDULE

<u>Lab. No.</u>	<u>Dates</u>	<u>Topics</u>
1.	18, 19 Jan.	Laboratory orientation: protocols, preparing solutions and making dilutions, pipeting, pipeting devices, and safety precautions
2.	25, 26 Jan.	Spectrophotometry: Beer's Law, use of the B&L Spectronic 20 spectrophotometers Constructing absorption curve for DCPIP and anthocyanin pigments at different pH values
3.	1, 2 Feb.	Spectrophotometric assays for protein (Bradford and bicinchoninic acid methods)
4.	8,9 Feb.	Cell growth: growth kinetics in <i>Enterobacter aerogenes</i> 1. Determining generation time 2. Effects of temperature, chloramphenicol, peptone, and an amino acid analogue
5.	15, 16 Feb.	Enzyme assay: acid phosphatase 1. Effect of substrate concentration 2. Effect of phosphate ion
6.	22, 23 Feb.	Estimation of specific activity of extracted acid phosphatase
7.	1, 2 Mar.	Isolating an organelle: 1. Mitochondria from cauliflower florets 2. Enzyme assay for succinic dehydrogenase and/or malate dehydrogenase
Sat.	5 Mar. - Sun. 13 Mar.	SPRING RECESS
8.	15, 16 Mar.	Factors affecting membrane permeability

9. 22, 23 Mar. Tyrosinase/phenol oxidase
10. 29, 30 Mar. Hill reaction and the Emerson enhancement effect in isolated chloroplasts
11. 5, 6 Apr. Manometric measurement of respiratory activity using the Gilson respirometer
1. Effects of temperature, substrates, and inhibitors on yeast Cell suspensions (*Saccharomyces cerevisiae*)
 2. Effects of washing and aging in discs of storage tissue from potato tubers (*Solanum tuberosum*)
12. 12, 13 Apr. Effects of ultraviolet radiation on *Paramecium*, *Tetrahymena*, and *Euglena*
13. 19, 20 Apr. Open
- Fri. 22 Apr. - Mon. 25 Apr. **EASTER RECESS**
14. 26, 27 Apr. Effects of cycloheximide and colchicine on flagella regeneration in *Chlamydomonas reinhardtii* (or) review

SUMMARY OF DEADLINES

Wed.	2 Feb.	First lab quiz (No's. 1 and 2)
Fri.	11 Feb.	FIRST HOUR EXAM
Wed.	2 Mar.	Second lab quiz (No's. 3, 4, 5 and 6)
Mon.	21 Mar.	SECOND HOUR EXAM
Wed.	6 Apr.	Third lab quiz (No's 7, 8, 9 and 10)
Wed.	20 Apr.	THIRD HOUR EXAM

